



## Fabrication of porous PCL/elastin composite scaffolds for tissue engineering applications

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### ABSTRACT

We present the development of a technique that enables the fabrication of three-dimensional (3D) porous poly( $\epsilon$ -caprolactone) (PCL)/elastin composites. High pressure CO<sub>2</sub> was used as a foaming agent to create large pores in a PCL matrix and impregnate elastin into the 3D structure of the scaffold. The effects of process variables such as temperature, pressure, processing time, depressurization rate, and salt concentration on the characteristics of PCL scaffolds were determined. Scaffolds with average pore sizes of 540  $\mu$ m and porosity of 91% were produced using CO<sub>2</sub> at 65 bar, 70 °C, processing time of 1 h, depressurization rate of 15 bar/min, and addition of 30 wt% salt particles. The PCL/elastin composites were then prepared under different conditions: ambient pressure, vacuum, and high pressure CO<sub>2</sub>. The fabrication of composites under vacuum resulted in the formation of nonhomogenous scaffolds. However, uniform 3D composites were formed when using high pressure CO<sub>2</sub> at 37 °C and 60 bar.

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### 1. Introduction

Polymers are used for the fabrication of hydrogel scaffolds in tissue engineering. These polymeric scaffolds are intended to support biological function by promoting the adhesion, differentiation, and viability of cells [1] and also to provide sufficient mechanical strength for the formation of functional engineered tissue [2]. Extracellular matrix (ECM) proteins such as collagen and elastin interact with cells via cell surface receptors and regulate or direct cell function [3]. However, their utility as hydrogel scaffolds has been limited by their poor mechanical properties. Synthetic biodegradable polymers such as poly( $\epsilon$ -caprolactone) (PCL), unlike natural ECM components, do not have specific cell-binding sites but do have superior mechanical strength [4,5]. The fabrication of hybrid synthetic/natural scaffolds allows for the incorporation of a suitable balance of biological and mechanical properties.

#### 1.1. Fabrication of composite scaffolds

Various methods have been used to combine natural and synthetic polymers for tissue engineering applications. Hybrid PCL/collagen films were fabricated by impregnation of freeze-dried collagen films with a solution of PCL in dichloromethane

followed by evaporation of solvent [6]. The fabricated composite scaffolds had pore sizes ranging from 50 to 100  $\mu$ m and could support the growth of human osteoblasts [6]. Two-dimensional (2D) PCL/natural polymer composite films have been prepared by coating PCL films prepared by solvent casting with biomimetic ECM components such as fibrin, gelatin, and fibronectin [7]. The fabricated composites significantly promoted endothelial cells adhesion and proliferation compared to pure PCL film [7]. Chen et al. fabricated collagen/poly(D,L-lactide-co-glycolide) (PLGA) scaffolds by embedding collagen fibers within a PLGA matrix [8]. In this method, PLGA sponges were prepared by a solvent casting/particle leaching technique using NaCl particles as the porogen and chloroform as the solvent [8]. The sponges were then immersed in an acidic collagen solution under vacuum to fill the pores of PLGA with collagen [8]. The composites were freeze-dried and subsequently cross-linked with glutaraldehyde (GA) vapor [8]. The Young's Modulus of the fabricated composite was 1.23 MPa which was higher than that of either PLGA (0.7 MPa) or collagen (0.2 MPa) [8]. A problem with the use of organic solvent in these methods is that any residue left in the material may be cytotoxic [9]. Electrospinning using natural and synthetic polymers can give hybrid natural/synthetic scaffolds [10,11]. Electrospinning was used to fabricate hybrid scaffolds comprised of PCL and natural proteins such as collagen, elastin, and gelatin, by dissolving them in hexafluoro-2-propanol [12]. The addition of PCL had no significant impact on porosity, but increased the mechanical properties of composites compared to protein alone. Collagen/elastin/PCL scaffolds presented pore sizes ranging from 8 to 39  $\mu$ m and Young's Modulus between 25 MPa and 35 MPa

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[12]. Elastin can also be combined with other synthetic polymers such as PLGA to produce hybrid electrospun biomaterials with improved mechanical properties for vascular applications where the modulus must be greater than 500 kPa [11]. The low thickness of fabricated scaffolds and residual organic solvent (e.g. hexafluoro-2-propanol) are major issues involved with using electrospinning for the formation of natural/synthetic composite scaffolds.

### 1.2. Fabrication of porosity

Cell adhesion and proliferation in scaffolds can be promoted by generating porosity within the 3D constructs [13]. Porosity is induced in polymeric matrices using a variety of methods including electrospinning, freeze-drying, and solvent casting/salt leaching [2]. However, the disadvantages of these techniques include the use of toxic organic solvent, formation of thin 2D structures, non-homogenous and limited porosity, irregularly shaped pores, and insufficient pore interconnectivity [2].

Gas foaming process using high pressure CO<sub>2</sub> has been widely employed to eliminate the problems associated with the use of these conventional methods for porosity generation. Porous structures of amorphous or semi-crystalline hydrophobic polymers such as poly(lactic acid) (PLA), PLGA, PCL, poly(methyl methacrylate) (PMMA) and polystyrene have been obtained using gas foaming technique [14–17]. There are three basic steps in this process: (a) polymer plasticization due to CO<sub>2</sub> diffusion into the polymer matrix with increasing pressure, (b) nucleation of gas bubbles as a result of depressurization and supersaturation, and (c) nucleation growth due to the gas diffusion from the surrounding polymer [16,18]. The formation of a non-porous external skin layer [19,20] and lack of interconnectivity between pores [21] due to the rheological and processing limitations are common issues in gas foaming technique. Gas foaming/salt leaching methods have been developed to address these issues [22,23]; for example Salerno et al. produced PCL foams with porosity in the range of 78–93% and pore sizes between 10 and 90 μm [23]. However, the gas foaming technique is not efficient for the creation of porosity in crystalline and hydrophilic polymers.

Recently we developed a technique to create porosity in composite tropoelastin/elastin hydrogels using high pressure CO<sub>2</sub> [24]. These composite hydrogels are formed in an aqueous phase without using any surfactant. The compressive modulus of the fabricated composite hydrogels increases 2-fold from 6.1 kPa to 11.8 kPa when high pressure CO<sub>2</sub> is used compared to hydrogels produced at 1 bar [24]. Further enhancement in mechanical properties of fabricated elastin-based hydrogels is required for engineering of load bearing tissues.

The aim of this study was to develop a rapid and solvent free process for the fabrication of homogenous PCL/elastin hybrid scaffolds suitable for tissue engineering applications, particularly bone and cartilage repair. Elastin is an insoluble ECM protein that provides various tissues in the body with the properties of extensibility and elastic recoil [25]. PCL is a biodegradable and biocompatible polyester with a low glass transition temperature (–60 °C) and melting point (60 °C), which make it easy to process [7]. PCL membranes elongate up to 1000% before break, which displays its high mechanical strength [4,5]. The results of our previous *in vitro* studies demonstrate that the addition of elastin to PCL dramatically promotes chondrocyte adhesion to PCL scaffolds [26]. In this study, the feasibility of using gas foaming/salt leaching technique for the creation of large pores (i.e. >500 μm) in PCL was assessed. The efficiency of high pressure CO<sub>2</sub> and vacuum for impregnation of elastin into the PCL porous structure to fabricate PCL/elastin composites was assessed by SEM, FTIR, weight gain elastin stability, and swelling ratio measurements.

## 2. Materials and methods

### 2.1. Materials

α-Elastin extracted from bovine ligament was purchased from Elastin Products Co. (MO, USA). PCL ( $M_w = 80$  kDa,  $T_m = 60$  °C,  $T_g = -60$  °C), GA and NaCl were purchased from Sigma–Aldrich. Food grade carbon dioxide (99.99% purity) was supplied by BOC. NaCl particles were ground and sieved to generate particles in the range of 100–700 μm.

### 2.2. Fabrication of PCL/elastin composite scaffolds

The schematic diagram for producing composite PCL/elastin scaffolds is illustrated in Fig. 1. The fabrication of hybrid scaffolds is characterized by four steps: (1) preparation of the PCL/NaCl blend by melt mixing; (2) gas foaming of the PCL/NaCl composite using dense gas CO<sub>2</sub>; (3) leaching out the salt particles from the PCL scaffold; (4) embedding elastin into the PCL scaffold and cross-linking under either high pressure CO<sub>2</sub>, atmospheric conditions or vacuum.

### 2.3. Formation of porous PCL scaffold

Experiments were conducted to determine the effects of gas foaming processing parameters on the pore characteristics of PCL. Process variables include saturation temperature ( $T_s$ ), saturation pressure ( $P_s$ ), soaking time ( $S_t$ ), depressurization rate (DPR), salt particle size and concentration. In each run, PCL was first melted at 60 °C and blended for at least 10 min with NaCl particles. The blend was then placed in a custom-made Teflon mold and cooled at 25 °C for 10 min to form disk-shaped samples ( $d = 5$  mm,  $h = 3$  mm).

A gas foaming process was then used to fabricate porous PCL scaffolds. The same experimental set-up as in our previous study was used for the gas foaming process [24]. In each run, a PCL/NaCl disk was placed inside a high pressure vessel (Thar, 100 ml view cell). The system was then pressurized with CO<sub>2</sub> to a predetermined  $P_s$  using a syringe pump (ISCO, Model 500D) and the pump was then run at constant pressure mode. The temperature was increased to the desired  $T_s$  using the Thar reactor temperature controller; the system was maintained at these conditions for a set period of  $S_t$ . The temperature was then gradually decreased to a foaming temperature of 34 °C at which point the inlet valve was closed and the system was depressurized at a predetermined DPR.

Fabricated PCL/NaCl samples were soaked in MilliQ water at room temperature for 24 h to leach out salt particles and then dried. Salt leaching ratio ( $\varphi$ ) measurements were used to determine the amount of NaCl leached out from the PCL scaffolds using the following equation:

$$\varphi = \frac{(W_b - W_a)/W_b}{C_{\text{NaCl}}} \times 100$$

where  $W_b$  is the weight of the PCL/NaCl sample before salt leaching,  $W_a$  is the weight of the PCL scaffold, and  $C_{\text{NaCl}}$  is the initial concentration of NaCl in PCL/NaCl blend.

#### 2.3.1. Fabrication of composite PCL/elastin scaffolds

Porous PCL scaffolds were soaked in an aqueous solution containing 5% (w/v) elastin and 0.25% (v/v) GA at 37 °C to fabricate composite scaffolds. Three different operating conditions comprising atmospheric pressure, vacuum, and high pressure CO<sub>2</sub> were used for embedding and cross-linking of elastin into the 3D structure of PCL. The cross-linking reaction was conducted for a period of 24 h when atmospheric pressure and vacuum (–1 MPa) were used to form the composites. The fabricated constructs were then washed repeatedly in PBS (10 mM phosphate, 150 mM NaCl; pH 7.4), and then placed in 100 mM Tris in PBS for 1 h to quench the

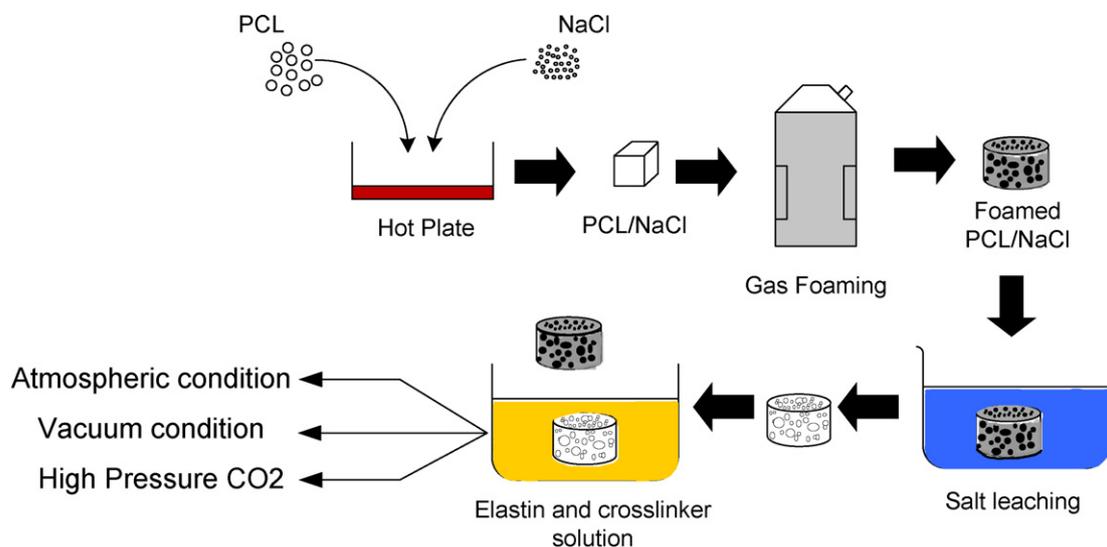


Fig. 1. Schematic diagram for the production of PCL/elastin composite scaffolds.

cross-linking reaction. After Tris treatment, the composite scaffolds were washed twice and stored in PBS.

The gas foaming apparatus was used for the preparation of PCL/elastin hybrid scaffolds at high pressure. The PCL scaffold was placed in a Teflon mold inside a high pressure vessel; elastin solution containing GA was then injected into the mold. After the vessel was sealed and equilibrated at 37 °C, the system was pressurized with CO<sub>2</sub> to 60 bar, isolated and maintained under these conditions for 1 h. The system was then depressurized at 15 bar/min and the sample was collected. The fabricated composite scaffold then quenched with Tris as described above.

Gravimetric analysis was used to determine the amount of elastin embedded in the 3D PCL scaffolds using the following equation:

$$\text{Weight gain} = \frac{W_{\text{Composite}} - W_{\text{PCL}}}{W_{\text{PCL}}} \times 100$$

where  $W_{\text{composite}}$  is the weight of the PCL/elastin composite scaffold and  $W_{\text{PCL}}$  is the weight of PCL scaffold.

### 2.3.2. Scanning electron microscopy (SEM)

SEM images of samples were obtained using a Zeiss Qemscan at 15 kV to determine the pore characteristics of the fabricated PCL scaffolds and to examine the penetration of elastin into the 3D structures of PCL matrices. Lyophilized constructs were mounted on aluminum stubs using conductive carbon paint then gold coated prior to SEM analysis.

### 2.4. Average pore size and porosity calculation

Image J software was used to calculate the equivalent circle diameter (ECD) of the pores using SEM images. For each sample several SEM images were taken. The sizes of at least 100 pores were measured using the Image J Software and the average pore sizes was then calculated.

Porosity of the scaffolds was calculated using the following equation:

$$\text{Porosity} = \frac{A_p}{A_T}$$

where  $A_p$  is total area of pores in each cross section and  $A_T$  is the total area of each cross section. Image J Software was used to calculate  $A_p$  and  $A_T$ . For each sample at least ten SEM images were

analyzed and the average porosity for each group of images was measured.

### 2.5. Fourier transform infrared (FTIR) spectrometer

FTIR analysis was used to qualitatively characterize the functional groups of elastin and PCL, and to confirm the 3D penetration of elastin into the PCL scaffolds. FTIR spectra were collected at the resolution of 2 cm<sup>-1</sup> and signal average of 32 scans in each interferogram over the range of 1900–1400 cm<sup>-1</sup> using a Varian 660 IR FTIR spectrometer. Composite scaffolds with thickness of 3 mm were used for FTIR analysis. The depth of elastin penetration into the PCL scaffolds was evaluated by performing FTIR analysis on the top surface and two layers manually cut from within the composites (i.e. 1 mm and 2 mm below the surface).

### 2.6. Water uptake properties

The water uptake ratios of PCL scaffold and PCL/elastin composites produced under high pressure CO<sub>2</sub> and vacuum were measured at 37 °C in PBS solution. The scaffolds were lyophilized to calculate their dried weight. The samples were then soaked in 10 ml PBS for 24 h. The excess liquid was removed from the samples and the water uptake was calculated based on a ratio of the increase in mass to that of the dry sample. The reported data at each condition was the average measurement for at least three scaffolds.

### 2.7. Elastin retention/stability ratio measurement

The stability of embedded crosslinked elastin within the 3D structures of composites was evaluated by immersing PCL/elastin composites fabricated under high pressure CO<sub>2</sub> in PBS solution at 37 °C for up to 7 days. The composites were taken out of the PBS solution at time intervals of 1, 2 and 7 days, washed with distilled water, freeze-dried and weighed. Elastin retention ratio measurement was used to determine the amount of elastin remaining within the 3D structures of composite scaffolds after soaking in PBS solution at different time intervals, using the following equation:

$$\text{Elastin retention ratio} = \frac{(W_{\text{composite}} - W_{\text{PCL}})_b}{(W_{\text{composite}} - W_{\text{PCL}})_a} \times 100$$

$(W_{\text{composite}} - W_{\text{PCL}})_a$  and  $(W_{\text{composite}} - W_{\text{PCL}})_b$  represent the weight of the elastin within the PCL scaffold after and before immersion in PBS solution for a given time, respectively.

### 2.8. Statistical analysis

Data is reported as mean  $\pm$  STD. One-way analysis of variance (ANOVA) with Bonferroni post hoc tests for multiple comparisons was performed using SPSS software for Windows version 18.0.1. Statistical significance was accepted at  $p < 0.05$  and indicated in the figures as  $*p < 0.05$ ,  $**p < 0.01$ , and  $***p < 0.001$ .

## 3. Results and discussion

The objective of this study was to create PCL/elastin composites with a large average pore size and a high degree of pore interconnectivity that could potentially be used for load-bearing tissue engineering applications including cartilage replacement. Chondrocyte proliferation and ECM production in gelatin scaffolds can be seen when the pore sizes are in the range of 250–500  $\mu\text{m}$  [13]. On this basis, PCL scaffolds were fabricated with pore sizes larger than 500  $\mu\text{m}$  so that following the subsequent elastin impregnation process, the PCL/elastin composites would retain pore sizes suitable for cellular penetration and growth.

Preliminary results for gas foaming neat PCL demonstrated that the process was not efficient for creating large interconnected pores. A skin layer was formed on the top surface of scaffold, when the gas foaming process was conducted at 65 bar, 70  $^{\circ}\text{C}$  for 1 h, and DPR of 15 bar/min; at these conditions the pore size was less than 150  $\mu\text{m}$  throughout the cross-section of sample. This result was in agreement with previous studies where PCL scaffolds with average pore sizes of 150  $\mu\text{m}$  were obtained at 250 bar, 40  $^{\circ}\text{C}$ , and DPR of 20 bar/min [27]. It has been demonstrated that the addition of NaCl particles to polymer as a hardening agent can increase pore-wall opening during foaming and enhance pore interconnectivity [23]. A gas foaming/salt leaching process was therefore investigated as a means of fabricating large interconnected pores in PCL scaffolds.

Gas foaming process parameters, NaCl particle size and concentration were manipulated to tailor the pore characteristics of PCL scaffolds.

### 3.1. Effect of salt

The effects of NaCl particle size (i.e. 100–300  $\mu\text{m}$ , 300–500  $\mu\text{m}$ , and 500–700  $\mu\text{m}$ ) and concentrations (30 wt%, 60 wt%) on pore characteristics and the leaching ratio ( $\varphi$ ) were determined. For this investigation, gas foaming process was conducted at 65 bar, 70  $^{\circ}\text{C}$ , using  $S_f$  of 1 h and DPR of 15 bar/min. A salt concentration of 30 wt% was used to determine the effect of NaCl particle sizes on pore morphology of fabricated PCL scaffolds. There was a correlation between the pore size of fabricated scaffold and salt particle size; as shown in Figs. 2 and 3A, increasing the salt particles from 100–300  $\mu\text{m}$  to 300–500  $\mu\text{m}$  significantly enhanced the pore sizes of PCL scaffolds from  $175 \pm 20 \mu\text{m}$  to  $390 \pm 15 \mu\text{m}$  ( $p < 0.01$ ). The highest average pore size of  $540 \pm 18 \mu\text{m}$  and porosity of  $91.1 \pm 1.2\%$  were obtained when porogen particle sizes of 500–700  $\mu\text{m}$  were used Fig. 3. The normalized PCL pore size distributions using various ranges of salt particle sizes are shown in Fig. 4. As shown in Fig. 4A, using a particle size range of 100–300  $\mu\text{m}$  resulted in generating pores with average size less than 400  $\mu\text{m}$ . The percentage of pores larger than 400  $\mu\text{m}$  approached 45% and 70% by increasing the porogen particle size to 300–500  $\mu\text{m}$  and 500–700  $\mu\text{m}$ , respectively (Fig. 4B and C).

Salt particle sizes in the range of 500–700  $\mu\text{m}$  were used to determine the effect of NaCl concentrations. As indicated in Figs. 2 and 3A, the pore sizes of PCL scaffolds significantly increased from  $310 \pm 15 \mu\text{m}$  to  $540 \pm 18 \mu\text{m}$  by decreasing the NaCl concentration from 60 wt% to 30 wt%, respectively ( $p < 0.001$ ). Increasing salt concentration from 30 wt% to 60 wt% significantly decreased the porosity of the fabricated scaffold from  $91.1 \pm 1.2\%$  to  $50 \pm 1.2\%$  ( $p < 0.001$ ), as shown in Fig. 3B. In addition,  $\varphi$  was enhanced from 70% to 100%, when the salt concentration was reduced from 60 wt% to 30 wt%. Salerno et al. observed a similar behavior for the effect of salt concentration on pore characteristics; PCL scaffolds with

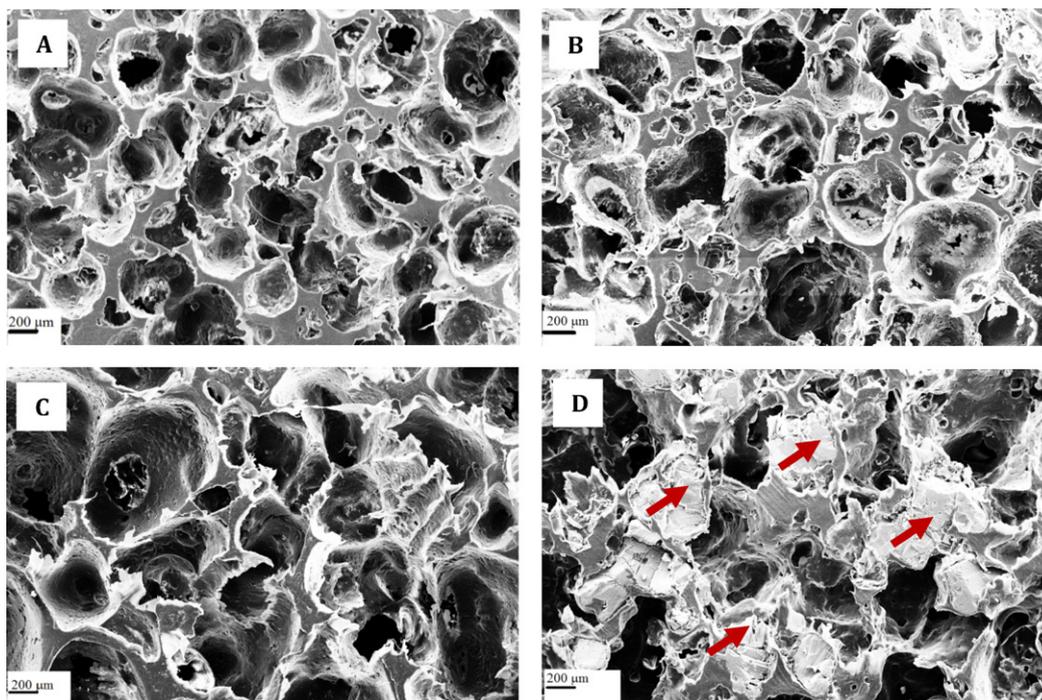


Fig. 2. SEM images of PCL scaffolds fabricated by the gas foaming/salt leaching process, using salt particle sizes of 100–500  $\mu\text{m}$  (A), 300–500  $\mu\text{m}$  (B), and 500–700  $\mu\text{m}$  (C and D). (Salt concentrations of 30 wt% were used in (A–C), and 60 wt% in (D); arrows show salt particles within pores of PCL scaffold.)

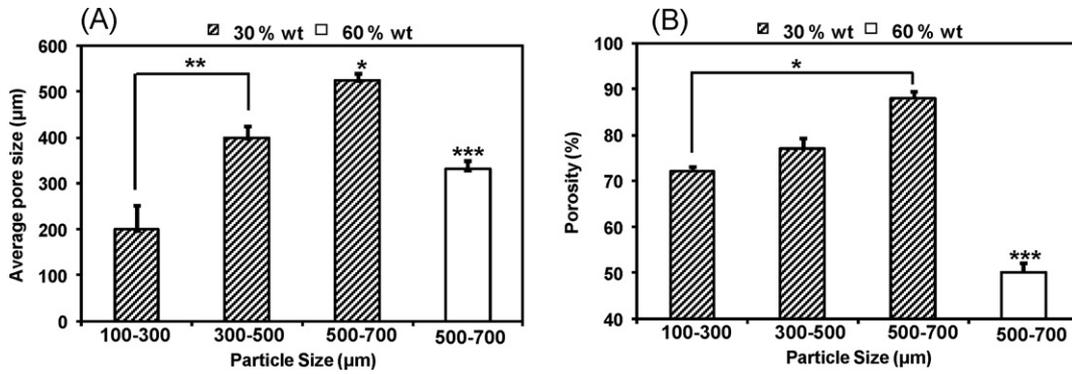


Fig. 3. Effect of salt particle size and concentration on the pore sizes (A) and porosity (B) of PCL scaffolds fabricated by the gas foaming/salt leaching process.

average pore size of 100 µm and 87% pore interconnectivity were produced using CO<sub>2</sub> at 65 bar, 70 °C for 3 h, and 30 wt% salt concentration [23]. These pore characteristics are not suitable for cartilage tissue engineering where pore sizes in the range of 250–500 µm are required [13].

Reduction in the pore sizes of fabricated scaffold, using 60 wt% NaCl, might be due to an increase in the stiffness of the PCL/NaCl blend, which resulted in limited CO<sub>2</sub> diffusion. Consequently, the porosity of scaffolds was decreased from 91.1 ± 1.2% to 50 ± 1.2%, by increasing the NaCl concentration from 30 wt% to 60 wt%. Salt particles were entrapped inside the structure of PCL scaffolds fabricated using 60 wt% NaCl, as shown in Fig. 2D. At this salt concentration, pores were mainly formed by gas foaming (<150 µm) and less pores were formed by salt leaching. Therefore, the average pore size of scaffold fabricated with 60 wt% NaCl was 320 µm, which was lower than the size of salt particle (500–700 µm) that were used.

It is critical to create highly interconnected pores with average size above 500 µm in PCL scaffold to facilitate both elastin penetration and cell proliferation in 3D structure. The presence of pore interconnectivity and large pore sizes can minimize surface

tension and facilitate elastin penetration into the porous structure of PCL. In this study, porogen concentration of 30 wt% and particle size in the range of 500–700 µm were used to fabricate PCL scaffolds.

### 3.2. Effect of gas foaming

Manipulation of processing parameters including  $P_s$ ,  $T_s$ , DPR, and  $S_t$  during gas foaming allow for control over the porous architecture of fabricated PCL scaffolds (Table 1). Preliminary results demonstrated that a non-uniform porous PCL scaffold was produced when the depressurization stage was conducted at temperatures above 34 °C. Di Maio et al. also reported that using foaming temperatures above 34 °C resulted in the formation of non-homogeneous PCL structures consisting of few large pores [28]. The formation of non-uniform pores in PCL structures can be due to PCL melting at high pressures and its inability to crystallize. Coalescence phenomena are likely to occur because of the low viscosity of PCL, which would not permit a complete stabilization of the cellular structure. Therefore, in this study a foaming

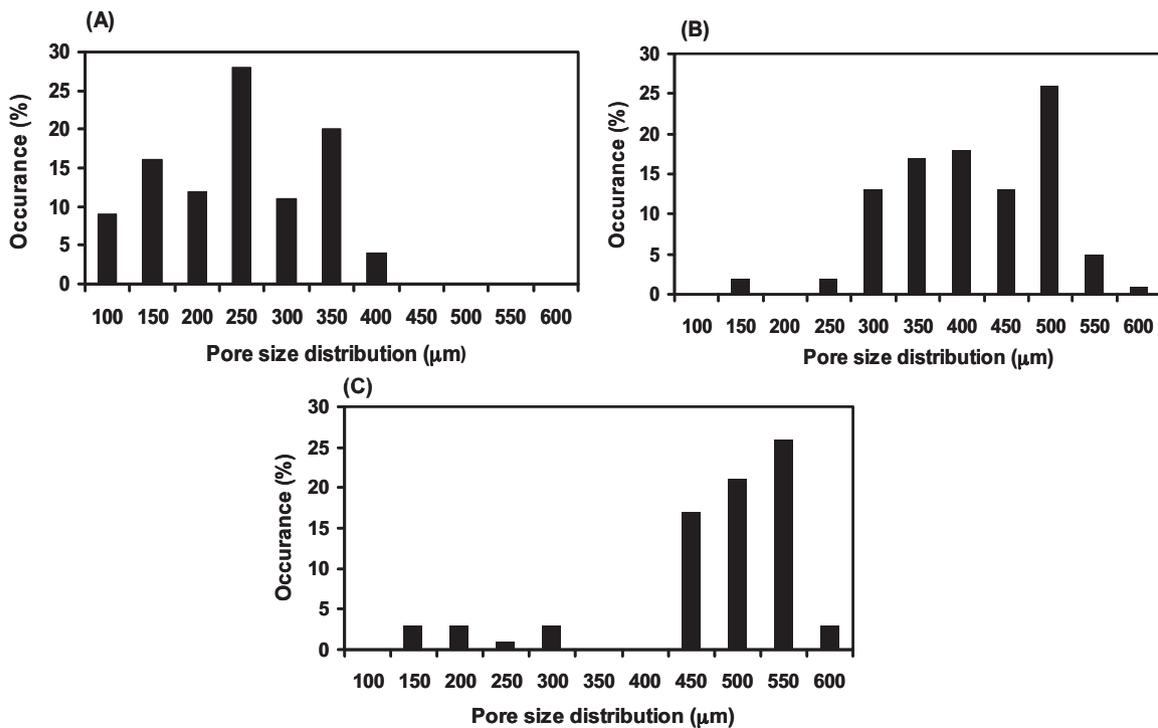


Fig. 4. Normalized pore size distribution of the PCL scaffolds fabricated using NaCl particles sizes of 100–300 µm (A), 300–500 µm (B), and 500–700 µm (C). (A salt concentration of 30 wt% was used.)

**Table 1**  
Effect of gas foaming parameters on leaching ratio, average pore sizes and porosity of PCL scaffold produced by using gas foaming/salt leaching process.

$P_s$ (bar)	$T_s$ ( $^{\circ}$ C)	$S_t$ (h)	DPR (bar/min)	$\varphi$ (%)	Average pore size ( $\mu$ m)	Porosity (%)
65	34	1	15	33	$210 \pm 20$	$23.1 \pm 3.2$
65	70	1	15	100	$540 \pm 18$	$91.1 \pm 1.2$
65	100	1	15	100	$530 \pm 21$	$89.2 \pm 3.3$
30	70	1	15	80	$320 \pm 15$	$51.9 \pm 4.5$
100	70	1	15	70	$310 \pm 15$	$64.2 \pm 2.1$
65	70	2	15	100	$545 \pm 10$	$91.2 \pm 2.9$
65	70	1	400	97	$158 \pm 10$	$60.1 \pm 1.1$

temperature of  $34^{\circ}$ C was used to investigate the effects of processing parameters on the pore morphology of fabricated PCL scaffolds.

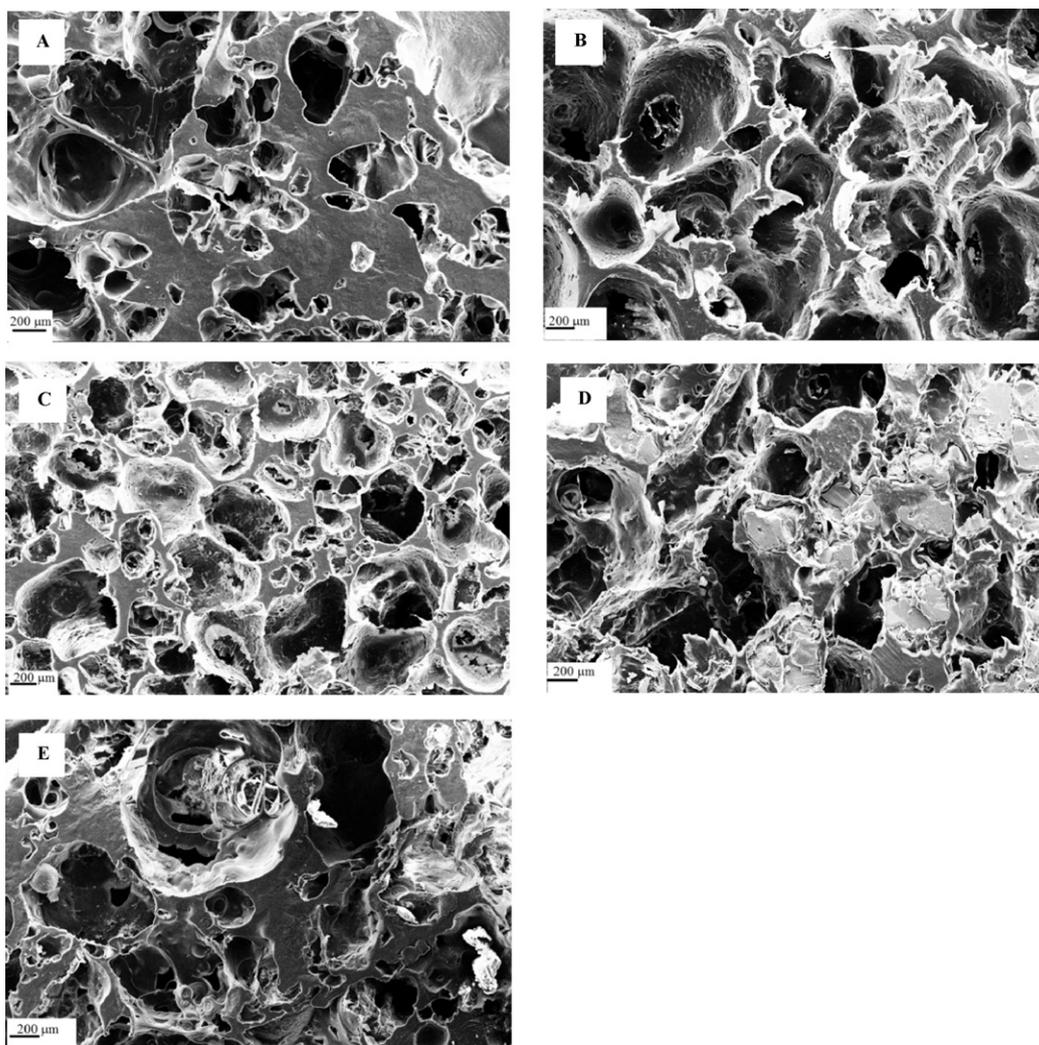
The effects of the gas foaming process variables including  $P_s$ ,  $T_s$ ,  $S_t$ , and DPR on the pore characteristics of PCL and  $\varphi$  were determined.  $T_s$  was varied between  $34^{\circ}$ C and  $100^{\circ}$ C and  $P_s$  in the range of 30–100 bar to investigate the effect of  $T_s$  and  $P_s$ . Two different rates of depressurization including 15 bar/min and 400 bar/min, and  $S_t$  of 1 h and 2 h were also used to investigate the effect of DPR and  $S_t$  on the pore size of PCL scaffolds, respectively.

### 3.3. Effect of saturation temperature ( $T_s$ )

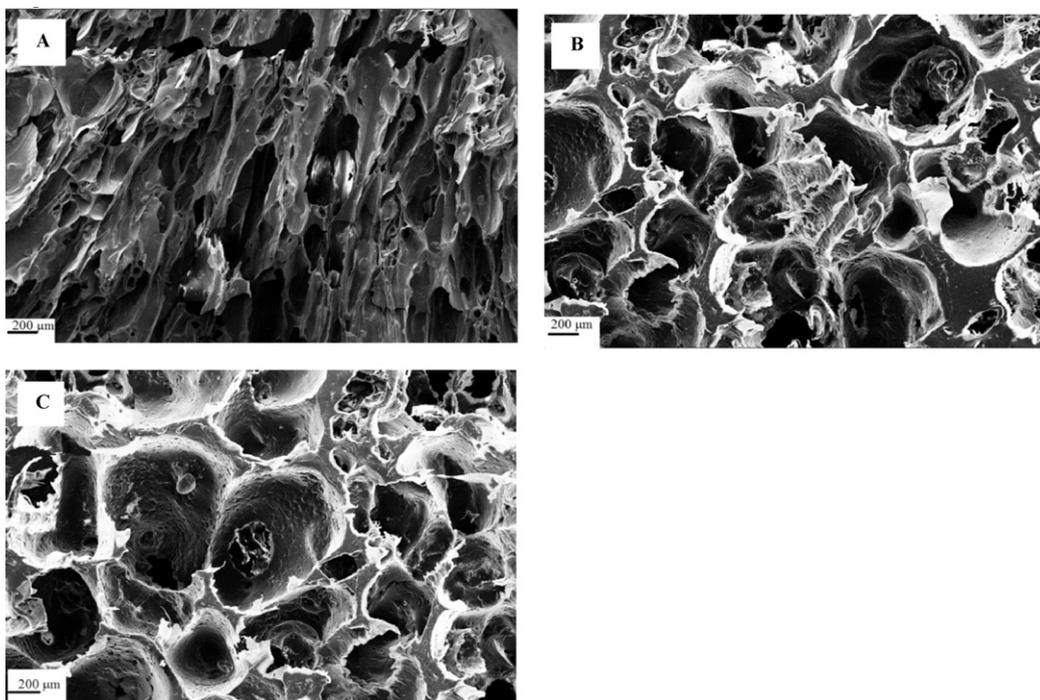
The effect of  $T_s$  on the pore morphology of fabricated PCL scaffolds was assessed at 65 bar for 1 h and using DPR of 15 bar/min. As

shown in Fig. 5 and Table 1, the average pore sizes of PCL scaffolds were significantly enhanced from  $210 \pm 20 \mu$ m to  $540 \pm 18 \mu$ m, when  $T_s$  was increased from  $34^{\circ}$ C to  $70^{\circ}$ C ( $p < 0.01$ ). In addition, only 33% of salt particles leached out when  $T_s$  of  $34^{\circ}$ C was used, confirming lower pore interconnectivity. In comparison, at  $T_s$  of  $70^{\circ}$ C complete salt removal could be obtained (Table 1). These results were in agreement with decreasing the porosity of scaffolds fabricated at lower saturation temperature,  $23.1 \pm 3.2\%$  compared to  $91.1 \pm 1.2\%$ . An elevation of  $T_s$  from  $70^{\circ}$ C to  $100^{\circ}$ C had negligible effect on the pore sizes and porosity of fabricated scaffolds; this can be explained by the effect of  $\text{CO}_2$  pressure on PCL melting point depression and also variation of the  $\text{CO}_2$  density at these two temperatures.

Increasing  $T_s$  reduces the viscosity of PCL, which results in the diffusion of  $\text{CO}_2$  within the polymer matrices and formation of scaf-



**Fig. 5.** SEM images of PCL scaffolds fabricated using the gas foaming/salt leaching process with a  $T_s$  of  $34^{\circ}$ C and 65 bar (A),  $70^{\circ}$ C and 65 bar, (B),  $100^{\circ}$ C and 65 bar (C),  $70^{\circ}$ C and 30 bar (D), and  $70^{\circ}$ C and 100 bar (E) (DPR of 15 bar/min, and  $S_t$  of 1 h were used).



**Fig. 6.** SEM images of PCL scaffold fabricated by the gas foaming/salt leaching process using DPR of 400 bar/min and  $S_t$  of 1 h (A), DPR of 15 bar/min and  $S_t$  of 2 h (B), and DPR of 15 bar/min and  $S_t$  of 1 h, ( $P_s$  of 65 bar and  $T_s$  of 70 °C were used).

folds with larger pores [15,29]. The PCL melting point is decreased from 60 °C to 46 °C when the CO<sub>2</sub> pressure is increased from 1 bar to 65 bar [30]; therefore, in this study PCL was melted at  $T_s$  above 46 °C and pressures higher than 60 bar. NIST REFPROP software was used to determine CO<sub>2</sub> density at various operating conditions. The density of CO<sub>2</sub> slightly decreased from 150 kg/m<sup>3</sup> to 145 kg/m<sup>3</sup> at 65 bar, when the temperature was increased from 70 °C to 100 °C. Therefore, the amount of CO<sub>2</sub> dissolved in molten PCL did not significantly change by raising temperature. It can be concluded that  $T_s$  of 70 °C was adequate for PCL gas foaming/salt leaching process.

#### 3.4. Effect of saturation pressure ( $P_s$ )

The  $P_s$  may have a significant impact on pore characteristics of PCL scaffolds. Previous studies demonstrated that no porosity is formed in PCL when it is pressurized with CO<sub>2</sub> at 70 °C and 30 bar [28]. However, increasing the processing pressure to 59 bar results in the formation of PCL scaffolds with average pore sizes of 250 μm [28]. In contrast, Tai et al. showed that the pore sizes of PLGA scaffolds fabricated by a gas foaming process are reduced when the processing pressure is increased from 60 bar to 150 bar [31]. In this study, gas foaming was conducted at  $T_s = 70$  °C, DPR = 15 bar/min, and  $S_t = 1$  h to investigate the effect of  $P_s$  on  $\phi$  and pore sizes of PCL scaffold (Table 1). As shown in Fig. 5, increasing  $P_s$  from 30 bar to 65 bar significantly increased the pore sizes of PCL from  $320 \pm 15$  μm to  $540 \pm 18$  μm ( $p < 0.01$ ). The effect of pressure on pore sizes can be explained by polymer plasticization and increasing CO<sub>2</sub> solubility in the molten phase at elevated pressures. A further increase of  $P_s$  to 100 bar reduced the average pore size of fabricated scaffolds to  $310 \pm 15$  μm; increasing the amount of CO<sub>2</sub> in the polymer phase increased gas nucleation density during the depressurization stage, resulting in the production of a large number of smaller pores [15]. Furthermore, it has been reported that the melting temperature of PCL decreases from 46 °C to 26 °C, when CO<sub>2</sub> pressure is increases from 65 bar to 100 bar [30]. Therefore, at

a foaming temperature of 34 °C and 65 bar, the PCL was not in a molten phase and solidified around the generated gas bubbles during depressurization; this resulted in the formation of large pores. However, at 34 °C and 100 bar; PCL was maintained in a molten phase during the depressurization stage which led to the generation of smaller pores.

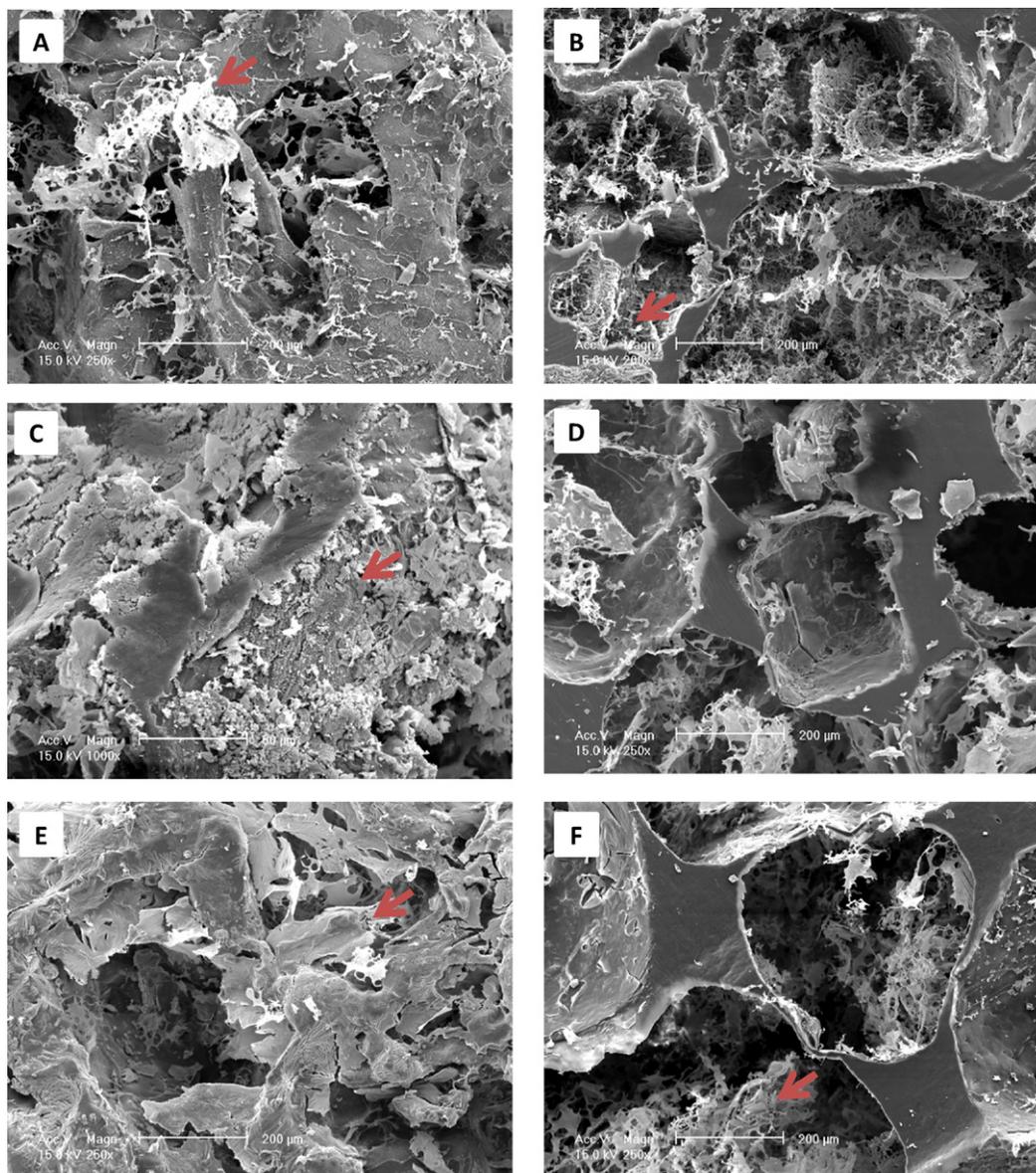
As shown in Table 1, salt particles had completely leached out when using a  $P_s$  of 65 bar; however,  $\phi$  was 80% and 70%, when the PCL was pressurized to 30 bar and 100 bar, respectively. Moreover, highest porosity,  $91.1 \pm 1.2\%$  was achieved when a saturation pressure of 65 bar was selected, as shown in Table 1. Therefore, a  $P_s$  of 65 bar was selected to fabricate PCL scaffold.

#### 3.5. Effect of depressurization rates (DPR)

The effect of DPR on pore morphology of PCL scaffolds was investigated by performing gas foaming at  $P_s = 65$  bar,  $T_s = 70$  °C, and  $S_t = 1$  h. As shown in Fig. 6, fast depressurization (400 bar/min) resulted in the formation of non-homogenous PCL scaffolds with average pore sizes of  $158.1 \pm 10$  μm. The pore sizes of fabricated PCL scaffolds were increased to  $540 \pm 18$  μm, when the DPR was decreased to 15 bar/min (Table 1). Tai et al. also reported the formation of non-homogenous structure of PLA scaffold produced at 35 °C, 230 bar for 1 h using a relatively fast depressurization of 115 bar/min; however, decreasing the DPR to 2 bar/min resulted in the formation of uniform PLA scaffolds with larger pores [31]. Lowering DPR can provide extra time for the gas nuclei to grow and generate larger pores [15,31].

#### 3.6. Effect of soaking time ( $S_t$ )

The effect of  $S_t$  on the pore morphology of fabricated PCL scaffolds was assessed at 65 bar, 70 °C for, and using DPR of 15 bar/min. As shown in Table 1 and Fig. 6,  $S_t$  had a negligible effect on the pore sizes and porosity of PCL scaffolds; increasing  $S_t$  from 1 h to



**Fig. 7.** SEM images of PCL/elastin composites fabricated using high pressure CO<sub>2</sub> (A and B), atmospheric pressure (C and D), and vacuum (E and F). Top surfaces are shown in A, C, and E; cross-sections in B, D, and F (arrows show the cross-linked elastin within the PCL scaffold).

2 h slightly changed the average pore size from  $540 \pm 18 \mu\text{m}$  to  $545 \pm 5 \mu\text{m}$ , and the porosity of both samples porosity was approximately 91%. This indicates that CO<sub>2</sub> saturated the PCL phase within 1 h because of the high mass transfer properties of CO<sub>2</sub> and also the small size of sample used in this study. Consequently, a soaking time of 1 h was considered adequate for the generation of appropriate pore sizes in the PCL scaffolds.

The porosity of a scaffold greatly affects the cellular adhesion and proliferation [13]. A highly porous scaffold was obtained for polymers such as PCL, PLA, and PLGA by using gas foaming/salt leaching process [23,31]. This process was efficient to create highly interconnected porous structures with porosity above 70% for these polymers [31]. It was also reported that 87% porosity was achieved for a PCL scaffold using 30 wt% salt concentration, and gas foaming by CO<sub>2</sub> at 65 bar and 70 °C [23]. The porosity of the PCL scaffold fabricated in this study was 91%. Interconnecting porosity achieved by gas foaming salt/leaching for PCL was further evidenced by cell infiltration into the construct acquired for this PCL/elastin scaffold in our recent study [26].

### 3.7. Fabrication of PCL/elastin composites

PCL/elastin composites were prepared under three different conditions: atmospheric pressure, vacuum, and high pressure CO<sub>2</sub>. We have previously shown that an elastin hydrogel is formed within 24 h when cross-linked at 37 °C under atmospheric pressure. In contrast, the hydrogel can be fabricated in less than 1 h when the cross-linking reaction is performed under high pressure CO<sub>2</sub> [32]. Therefore, PCL scaffolds were soaked in elastin solution containing GA for 1 h at high pressure and 24 h under vacuum and atmospheric conditions to form composite constructs.

FTIR, SEM and gravimetric techniques were used to assess the efficiency of each process for the creation of homogenous 3D hybrid scaffolds. The presence of cross-linked elastin on the top surface (Fig. 7A) and within the cross section (Fig. 7B) of PCL scaffolds fabricated under high pressure CO<sub>2</sub> confirmed that elastin uniformly penetrated within the 3D structure of PCL under these conditions. The composite samples comprised two pore size components: 500 μm-sized PCL components and the smaller pores of

elastin hydrogels, similar to those produced previously [32]. In contrast, immersing the PCL scaffold in an elastin solution under atmospheric conditions resulted in the formation of a layer of cross-linked elastin on the top surface of the PCL foam with no penetration into the 3D structure (Fig. 7C and D). This effect might be due to the surface tension of the water phase that impeded solution penetration into the pores of PCL. Atmospheric conditions are therefore only useful for the formation of an elastin surface coating of PCL scaffolds. Impregnation of PCL scaffolds with elastin under vacuum conditions for 24 h slightly improved the penetration of elastin into the PCL scaffold, but the elastin was not homogeneously distributed throughout the PCL structure as shown in Fig. 7E and F.

FTIR spectra of an elastin hydrogel showed two main peaks at  $1535\text{ cm}^{-1}$ , and  $1655\text{ cm}^{-1}$  corresponding to the amide II and amide I bands, respectively (Fig. 8). Similar peaks were observed for bovine elastin and *k*-elastin [33], human elastin [34], bovine tropoelastin [35],  $\alpha$ -elastin [36,37], two elastin-like poly(pentapeptides) [38], and synthetic elastin hydrogel [39]. The FTIR spectra of PCL showed a main peak at  $1725\text{ cm}^{-1}$  attributed to C=O stretching; this peak was also observed in previous studies for electrospun PCL samples [40]. The presence of peaks assigned to PCL and elastin on the top surface and 1 mm and 2 mm into the hybrid scaffold fabricated at high pressure  $\text{CO}_2$  confirmed the presence of elastin in all layers of composites (Fig. 8A). However, FTIR analysis on hybrid scaffolds produced under vacuum demonstrated elastin penetration up to approximately 1 mm into the PCL scaffold as shown in Fig. 8B. The results of FTIR and SEM analysis confirmed that high pressure  $\text{CO}_2$  was more efficient than the other techniques used in this study for the production of homogenous 3D hybrid PCL/elastin scaffolds.

The weight gain and water uptake of composites fabricated under high pressure  $\text{CO}_2$  and vacuum are shown in Table 2. Samples fabricated under vacuum exhibited lower mass gain of  $3 \pm 0.08\text{ wt}\%$  compared with those produced at high pressure  $\text{CO}_2$  ( $15 \pm 0.07\text{ wt}\%$ ). This was due to an increased introduction of elastin

**Table 2**

The weight gain and water uptake ratio of PCL/elastin composites fabricated by using high pressure  $\text{CO}_2$  and vacuum.

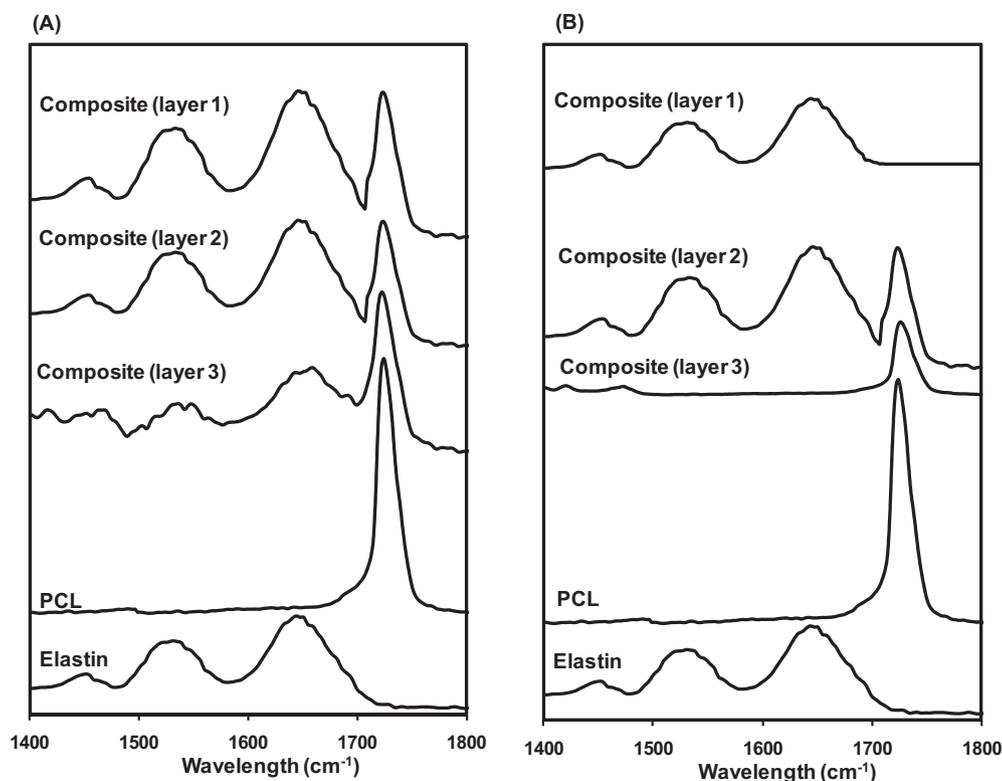
Composite scaffolds	Weight gain (wt%)	Water uptake ratio (g liquid/g polymer)
High pressure $\text{CO}_2$	$15 \pm 0.07$	$3.5 \pm 0.1$
Vacuum	$3 \pm 0.08$	$1.1 \pm 0.1$

into the pores of PCL scaffolds when high pressure  $\text{CO}_2$  was used for composite fabrication compared to vacuum.

The presence of elastin within the 3D PCL matrices imparted hydrophilicity to the composite scaffolds and affected their water uptake properties. As shown in Table 2, hybrid scaffolds fabricated under high pressure  $\text{CO}_2$  exhibited higher water uptake ratio of  $3.5 \pm 0.1\text{ g liquid/g polymer}$  compared with the one produced under vacuum conditions ( $1.1 \pm 0.1\text{ g liquid/g polymer}$ ). Generally, the water uptake ratios of composites fabricated under either high pressure  $\text{CO}_2$  or vacuum were higher than PCL scaffolds ( $0.03 \pm 0.005\text{ g liquid/g polymer}$ ).

The compressive modulus of fabricated hybrid scaffolds was higher than of pure elastin hydrogel and lower than of PCL scaffold [26]. The compressive modulus of PCL/elastin composites was  $1.30 \pm 0.07\text{ MPa}$ , which slightly exceeded the target value for cartilage repair,  $0.5\text{--}1\text{ MPa}$  [26].

The stability of adsorbed crosslinked elastin within the 3D structure of composites was evaluated by calculating the elastin retention ratio after soaking in PBS for 1, 2, and 7 days. As confirmed by FTIR and weight gain analyses, elastin diffusion throughout the 3D PCL scaffolds was higher for the PCL/elastin composites fabricated under high pressure  $\text{CO}_2$  compared to those produced by using vacuum. Therefore, elastin stability was only determined for the composites fabricated under high pressure  $\text{CO}_2$ . As shown in Fig. 9, elastin retention ratio diminished from  $86.4 \pm 2\text{ (w/w) \%}$  at day 1 to  $72.1 \pm 1\text{ (w/w) \%}$  and  $70.7 \pm 2\text{ (w/w) \%}$  at days 2 and 7 of



**Fig. 8.** Fourier transform infrared (FTIR) spectra for pure  $\alpha$ -elastin hydrogel, PCL, and composite PCL/ $\alpha$ -elastin scaffolds produced under high pressure  $\text{CO}_2$  (A), and vacuum (B) (Layer 1: top surface, Layer 2: 1 mm depth, Layer 3: 2 mm depth).

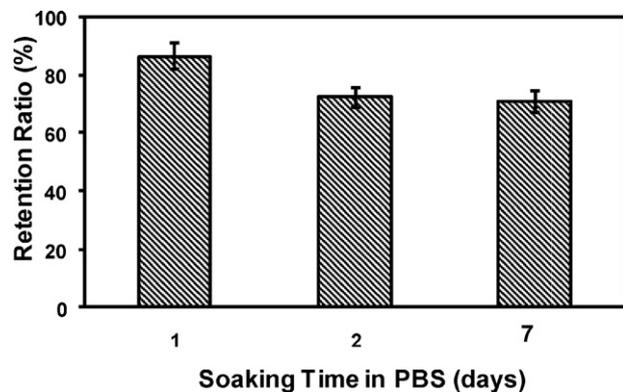


Fig. 9. Elastin retention in composite PCL/elastin scaffolds fabricated under high pressure CO<sub>2</sub> on days 1, 2, and 7 of immersion in PBS.

soaking, respectively. The decrease in retention ratio on day 2 of immersing in PBS may be due to the removal of excess uncross-linked elastin from the composites during washing step. However, there was no significant change in retention ratio of elastin after the second day of soaking. This confirmed the stability of cross-linked elastin within the pores of hybrid scaffolds fabricated under high pressure CO<sub>2</sub>.

The hybrid scaffold was comprised of large size pores of PCL construct (average size of 540 μm) and small size pores of elastin hydrogel (average size of 50 μm). The results of *in vitro* studies confirmed the proliferation of chondrocytes into the 3D structure of hybrid scaffold processed by dense gas CO<sub>2</sub> [26]. These results demonstrate that the pores in the composite construct were interconnected and nutrient and oxygen could penetrate through the 3D structure of composite scaffold.

Hybrid natural/synthetic scaffolds such as chitosan/PCL [41], collagen/PCL [6], chitosan/PLLA [42], and alginate/2-hydroxyethylmethacrylate (HEMA) [43] have been constructed for tissue engineering applications; generally, a porous structure of one polymer is immersed in a solution of the other polymer under vacuum to form these composites [6,41,42]. Mei et al. made PCL/chitosan scaffolds by soaking prefabricated PCL foams, using solvent casting/particle leaching, in a solution of chitosan under vacuum pressure. The weight gain of PCL/chitosan composite is 8.4 ± 0.1 wt% when 20 mg/ml of chitosan solution is used [41]. Porous alginate/HEMA hybrids can be made by supercritical CO<sub>2</sub> assisted impregnation of alginate foams with HEMA and a cross-linking agent [43] where the pore size and weight gains of fabricated composites are in the range of 21–105 μm and 40–170 wt%, respectively, depending on the HEMA and crosslinker concentrations. Jiao et al. produced a hybrid chitosan/PLLA scaffold by immersing PLLA scaffold, produced by a solvent casting/salt leaching process, into a 1.5 wt% chitosan solution using vacuum [43]. When the chitosan concentration is higher than 1.5 wt%, it is difficult for the solution to penetrate into the PLLA sponge due to its high viscosity [43]. In our study, the negligible diffusion of cross-linked elastin within 3D PCL scaffold fabricated under vacuum appears to be due to the high viscosity of the cross-linking elastin solution within 24 h of cross-linking time. However, elastin solution penetrated into 3D structure of porous PCL under high pressure and cross-linked. High pressure CO<sub>2</sub> was the driving force to overcome the surface tension and viscosity of elastin solution that can impede diffusion of solution into the porous structure. High pressure CO<sub>2</sub> allowed us to overcome limitations posed by more conventional methods (e.g. vacuum) such as long processing time (e.g. 24 h) [6,8], use of organic solvent [6,8,41], and the formation of 2D structure [6].

#### 4. Conclusions

Dense gas processes were successfully used to create both porous PCL scaffolds with a high degree of interconnectivity and homogenous 3D PCL/elastin hybrid scaffolds. Porous PCL scaffolds with average pore sizes larger than 500 μm and porosity of 91% were produced using a gas foaming/salt leaching process. The fabrication of hybrid scaffolds at atmospheric pressure was efficient for surface modification of PCL. Conducting the process under vacuum allowed for elastin penetration up to 1 mm into the samples which may be useful for the fabrication of thin hybrid scaffolds. However, the fabrication of PCL/elastin composites under high pressure CO<sub>2</sub> enabled elastin penetration throughout the 3D PCL scaffolds. The water uptake ratio of hybrid scaffolds fabricated using high pressure CO<sub>2</sub> was 3-fold higher than those produced under vacuum. Composites fabricated using high pressure CO<sub>2</sub> may be very useful for tissue engineering applications.

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#### References

- [1] D.V. Bax, D.R. McKenzie, A.S. Weiss, M.M.M. Bilek, The linker-free covalent attachment of collagen to plasma immersion ion implantation treated polytetrafluoroethylene and subsequent cell-binding activity, *Biomaterials* 31 (2009) 2526–2534.
- [2] N. Annabi, J.W. Nichol, X. Zhong, C. Ji, S. Koshy, A. Khademhosseini, F. Dehghani, Controlling the porosity and microarchitecture of hydrogels for tissue engineering, *Tissue Engineering: Part B* 16 (4) (2010) 371–380.
- [3] P.B. Malafaya, G.A. Silva, R.L. Reis, Natural-origin polymers as carriers and scaffolds for biomolecules and cell delivery in tissue engineering applications, *Advanced Drug Delivery Reviews* 59 (4–5) (2007) 207–233.
- [4] A. Sarasam, S.V. Madhally, Characterization of chitosan–polycaprolactone blends for tissue engineering applications, *Biomaterials* 26 (2005) 5500–5508.
- [5] L. Averousa, L. Moro, P. Dole, C. Fringant, Properties of thermoplastic blends: starch–polycaprolactone, *Polymer* 41 (2000) 4157–4167.
- [6] A.G.A. Coombes, E. Verderio, B. Shaw, X. Li, M. Griffin, S. Downes, Biocomposites of non-crosslinked natural and synthetic polymers, *Biomaterials* 23 (2002) 2113–2118.
- [7] D. Pankajakshan, K.V. Kalliyana, L.K. Krishnan, Vascular tissue generation in response to signaling molecules integrated with a novel poly(1,4-caprolactone)–fibrin hybrid scaffold, *J. Tissue Engineering and Regenerative Medicine* 1 (5) (2007) 389–397.
- [8] G. Chen, T. Ushida, T. Tateishi, A biodegradable hybrid sponge nested with collagen microsponges, *J. Biomedical Materials Research* 51 (2) (2000) 273–279.
- [9] E. Sachlos, J.T. Czernuszka, Making tissue engineering scaffolds work, *European Cells & Materials* 5 (2003) 29.
- [10] J. Stitzel, J. Liu, S.J. Lee, M. Komura, J. Berry, S. Soker, G. Lim, M. Van Dyke, R. Czerw, J.J. Yoo, A. Atala, Controlled fabrication of a biological vascular substitute, *Biomaterials* 27 (7) (2006) 1088–1094.
- [11] M.K.C. Ng, A.S. Weiss, S.G. Wise, Tropoelastin-based protoelastin biomaterials of high flexibility for use in vascular prosthetic applications, *WO 2008/037028*.
- [12] S. Heydarkhan-Hagvall, K. Schenke-Layland, A.P. Dhanasopon, F. Rofail, H. Smith, B.M. Wu, R. Shemin, R.E. Beygui, W.R. MacLellan, Three-dimensional electrospun ECM-based hybrid scaffolds for cardiovascular tissue engineering, *Biomaterials* 29 (19) (2008) 2907–2914.
- [13] S.-M. Lien, L.-Y. Ko, T.-J. Huang, Effect of pore size on ECM secretion and cell growth in gelatin scaffold for articular cartilage tissue engineering, *Acta Biomaterialia* 5 (2) (2009) 670–679.
- [14] Z.-M. Xu, X.-L. Jiang, T. Liu, G.-H. Hu, L. Zhao, Z.-N. Zhu, W.-K. Yuan, Foaming of polypropylene with supercritical carbon dioxide, *J. Supercritical Fluids* 41 (2) (2007) 299–310.
- [15] H. Tai, V.K. Popov, K.M. Shakesheff, S.M. Howdle, Putting the fizz into chemistry: applications of supercritical carbon dioxide in tissue engineering, drug delivery and synthesis of novel block copolymers, *Biochemical Society Transactions* 35 (3) (2007) 516–521.
- [16] J.J.A. Barry, M.M.C.G. Silva, V.K. Popov, K.M. Shakesheff, S.M. Howdle, Supercritical carbon dioxide: putting the fizz into biomaterials, *Philosophical Transactions Series A: Mathematical, Physical and Engineering Sciences* 364 (1838) (2006) 249–261.
- [17] J.J.A. Barry, M.M.C.G. Silva, S.H. Cartmell, R.E. Guldberg, C.A. Scotchford, S.M. Howdle, Porous methacrylate tissue engineering scaffolds: using carbon dioxide to control porosity and interconnectivity, *J. Materials Science* 41 (13) (2006) 4197–4204.

- [18] C.B. Park, D.F. Baldwin, N.P. Suh, Effect of the pressure drop rate on cell nucleation in continuous processing of microcellular polymers, *Polymer Engineering and Science* 35 (5) (1995) 432–440.
- [19] D.J. Mooney, D.F. Baldwin, N.P. Suh, J.P. Vacanti, R. Langer, Novel approach to fabricate porous sponges of poly(D,L-lactic-co-glycolic acid) without the use of organic solvents, *Biomaterials* 17 (14) (1996) 1417–1422.
- [20] S.K. Goel, E.J. Beckman, Generation of microcellular polymeric foams using supercritical carbon dioxide. II. Cell growth and skin formation, *Polymer Engineering and Science* 34 (14) (1994) 1148–1156.
- [21] D. Hutmacher, Scaffolds in tissue engineering bone and cartilage, *Biomaterials* 21 (2000) 2529–2543.
- [22] L.D. Harris, B.S. Kim, D.J. Mooney, Open pore biodegradable matrices formed with gas foaming, *J. Biomedical Materials Research* 42 (3) (1998) 396–402.
- [23] A. Salerno, S. Iannace, P.A. Netti, Open-pore biodegradable foams prepared via gas foaming and microparticle templating, *Macromolecular Biosciences* 8 (2008) 655–664.
- [24] N. Annabi, S.M. Mithieux, A.S. Weiss, F. Dehghani, Cross-linked open-pore elastic hydrogels based on tropoelastin, elastin and high pressure CO<sub>2</sub>, *Biomaterials* 31 (7) (2010) 1655–1665.
- [25] S.M. Mithieux, A.S. Weiss, Elastin, *Advances in Protein Chemistry* 70 (2005) 437–461.
- [26] N. Annabi, A. Fathi, S.M. Mithieux, P. Martens, A.S. Weiss, F. Dehghani, The effect of elastin on chondrocyte adhesion and proliferation on poly( $\epsilon$ -caprolactone) elastin composites, *Biomaterials* 32 (2011) 1517–1525.
- [27] S. Blacher, C. Calberg, G. Kerckhofs, A. Léonard, M. Wevers, R. Jérôme, J.-P. Pirard, The porous structure of biodegradable scaffolds obtained with supercritical CO<sub>2</sub> as foaming agent, *Studies in Surface Science and Catalysis* 160 (2007) 681–688.
- [28] E. Di Maio, G. Mensitieri, S. Iannace, L. Nicolais, W. Li, R.W. Flumerfelt, Structure optimization of polycaprolactone foams by using mixtures of CO<sub>2</sub> and N<sub>2</sub> as blowing agents, *Polymer Engineering and Science* 45 (3) (2005) 432–441.
- [29] K.A. Arora, A.J. Lesser, T.J. McCarthy, Preparation and characterization of microcellular polystyrene foams processed in supercritical carbon dioxide, *Macromolecules* 31 (14) (1998) 4614–4620.
- [30] Z. Lian, S. Epstein, C. Blenk, A. Shine, Carbon dioxide-induced melting point depression of biodegradable semicrystalline polymers, *J. Supercritical Fluids* 39 (2006) 107–117.
- [31] H. Tai, M.L. Mather, D. Howard, W. Wang, L.J. White, J.A. Crowe, S.P. Morgan, A. Chandra, D.J. Williams, S.M. Howdle, K.M. Shakesheff, Control of pore size and structure of tissue engineering scaffolds produced by supercritical fluid processing, *European Cells & Materials* 14 (2007) 64–77.
- [32] N. Annabi, S.M. Mithieux, A.S. Weiss, F. Dehghani, The fabrication of elastin-based hydrogels using high pressure CO<sub>2</sub>, *Biomaterials* 30 (1) (2009) 1–7.
- [33] L. Debelle, A.J.P. Alix, M.-P. Jacob, J.-P. Huvenne, M. Berjot, B. Sombret, P. Legrand, Bovine elastin and k-elastin secondary structure determination by optical spectroscopies, *J. Biological Chemistry* 270 (44) (1995) 26099–26103.
- [34] L. Debelle, A.J.P. Alix, S.M. Wei, M.-P. Jacob, J.-P. Huvenne, M. Berjot, P. Legrand, The secondary structure and architecture of human elastin, *European Journal of Biochemistry* 258 (2) (1998) 533–539.
- [35] L. Debelle, A.J.P. Alix, Optical spectroscopic determination of bovine tropoelastin molecular model, *J. Molecular Structure* 348 (1995) 321–324.
- [36] M. Mammi, L. Gotte, G. Pezzin, Evidence for order in the structure of alpha-elastin, *Nature* 220 (5165) (1968) 371–373.
- [37] F. Dehghani, N. Annabi, P. Valtchev, S.M. Mithieux, A.S. Weiss, S.G. Kazarian, F.H. Tay, Effect of dense gas CO<sub>2</sub> on the coacervation of elastin, *Biomacromolecules* 9 (4) (2008) 1100–1105.
- [38] P. Schmidt, J. Dybal, J.C. Rodriguez-Cabello, V. Reboto, Role of water in structural changes of poly(AVGVP) and poly(GVGVP) studied by FTIR and Raman spectroscopy and ab initio calculations, *Biomacromolecules* 6 (2) (2005) 697–706.
- [39] S.M. Mithieux, J.E.J. Rasko, A.S. Weiss, Synthetic elastin hydrogels derived from massive elastic assemblies of self-organized human protein monomers, *Biomaterials* 25 (20) (2004) 4921–4927.
- [40] R. Nirmala, K.T. Nam, D.K. Park, B. Woo-il, R. Navamathavan, H.Y. Kim, Structural, thermal, mechanical and bioactivity evaluation of silver-loaded bovine bone hydroxyapatite grafted poly([epsilon]-caprolactone) nanofibers via electrospinning, *Surface and Coating Technology* 205 (1) (2010) 174–181.
- [41] N. Mei, G. Chen, P. Zhou, X. Chen, Z.-Z. Shao, L.-F. Pan, C.-G. Wu, Biocompatibility of poly( $\epsilon$ -caprolactone) scaffold modified by chitosan—the fibroblasts proliferation in vitro, *J. Biomaterials Applications* 19 (4) (2005) 323–339.
- [42] Y. Jiao, Z. Liu, C. Zhou, Fabrication and characterization of PLLA–chitosan hybrid scaffolds with improved cell compatibility, *J. Biomedical Materials Research, Part A* 80A (4) (2007) 820–825.
- [43] S. Partap, A.K. Hebb, I. ur Rehman, J.A. Darr, Formation of porous natural-synthetic polymer composites using emulsion templating and supercritical fluid assisted impregnation, *Polymer Bulletin* 58 (5–6) (2007) 849–860.